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⑤④ Immunosuppressant factor.

⑤⑦ An immunosuppressant factor, a neuroblast growth  
inhibitory factor and an interleukin 1 like factor derived from  
human glioblastoma cells.

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Case 118-6323IMMUNOSUPPRESSANT FACTOR

The present invention is concerned with immunomodulatory substances.

It is more particularly concerned with an immunosuppressant factor capable of inhibiting T-cell mechanisms particularly those which are Interleukin 2 (IL-2) dependent.

5       The role of T-cells in cell-mediated immunity e.g. by cooperation with  $\beta$ -lymphocytes is well established and substances which are capable of inhibiting T-cell mechanisms involved in the generation of antibodies would be used to suppress or reduce the body's immune response. Such substances could thus be employed e.g. in connection with transplants to  
10 prevent rejection and also in connection with the treatment of diseases characterised by an auto-immune response in the body.

We have now found that cultured human glioblastoma cells secrete a factor that inhibits interleukin 2 (IL-2) dependent T-cell mechanisms. This factor has an inhibitory effect on IL-2 induced proliferation of  
15 T-cell clones and on the induction of alloreactive cytotoxic T-cells in mixed lymphocyte cultures. It further inhibits the growth of neuroblasts but not of fibroblasts.

This factor can be obtained from the cystic fluid or SN of cultured human glioblastoma cell lines and has an approximate molecular weight  
20 of 97,000.

Cultured human glioblastoma cells are characterized by ultra-structural features, especially the presence of microfibrils, and by biochemical markers, such as glial fibrillary acidic protein (GFA) and S-100 protein (7, 14, 15). In addition human glioma cells express neu-  
25 roectodermal antigens shared with melanomas and neuroblastomas (8), as well as Ia-like antigens and the common acute lymphoblastic leukemia antigen (CALLA), an antigen expressed on lymphoid cells from patients with the common form of acute lymphoblastic leukemia (16, 17).

Two immunoregulatory mediators, a suppressor factor and a helper factor, were detected in the supernatant of glioblastoma cells. The helper factor shares the characteristics of macrophage-derived IL 1. The characterization of the factor as an IL 1-like mediator is based on the finding that the factor a) enhances the PHA-induced thymocyte proliferation, b) exhibits no IL 2 activity, but c) augments the IL 2 production by nitrogen-stimulated spleen cells, and d) has a m.w. of 22,000. In this context it is of interest that mouse astrocytes stimulated with lipopolysaccharide and rat C<sub>6</sub> glioma cells secrete analogous IL 1-like factors in vitro (9, 18, 19). Furthermore, rat astrocytes have been shown to present antigen to T-lymphocytes, e.g. to activate myelin basic protein-specific T lymphocyte lines (20). In addition, cultured rat astrocytes express Ia antigens upon interaction with T cells (20). The production of IL 1-like factors and the presentation of antigen by astrocytes may have a central role in the generation of immune responses in the brain. There is also recent evidence that the central nervous system is capable of mounting specific immune responses to antigens and tumor cells when introduced to the brain (21).

Analogous to astrocytes, glioblastoma cells, which may represent transformed astrocyte precursor cells, do produce IL 1-like factors in vitro. The effect of the IL 1-like helper factor, however, is overridden by the presently described inhibitory factor also elaborated by the tumor cells. This factor inhibits the proliferative response of T cells to Con A or to both PHA and IL 1/IL 2 standards. The factor also blocks the proliferation of an H-2-restricted, hapten-specific T cell line that normally grows in the presence of IL 2 and haptenated irradiated spleen cells. The glioblastoma-derived suppressor factor has no effect on IL 2 production by Con A-stimulated spleen cells and does not influence the growth of a thymoma cell line (EL 4 cells), which proliferate independently of IL 2. However, the factor abrogates the IL 2-induced proliferation of a cloned T cell line. IL 2 does not substitute for the inhibitor effect of the supernatant containing the factor on the development of CTL. Taken together, these results indicate that the factor interferes with

terminal events in the T cell activation cascade. The hypothesis that this glioblastoma-derived suppressor factor influences proliferative steps not necessarily involving direct IL 2 effects on T cells is based on the finding that the 97,000-m.w. factor does also inhibit the growth of neuroblasts, which grow independently of IL 2.

Recently, the presence of immunosuppressive factors in the conditioned medium of various human tumor cells was investigated (22). Out of 12 different tumors (three melanoma, two sarcoma, and seven carcinoma) the SN of only three tumors produced greater than 30% inhibition of PHA-induced blastogenesis. However, as shown by dilution experiments the inhibitory effect was only visible up to a  $1/10$  dilution of the SN; higher dilutions had no effect (22). When using the conditions to produce and to test the glioblastoma SN, the SN of two neuroblastoma, a melanoma, and a rhabdomyosarcoma had no effect on responsiveness of thymocytes to Con A. This supports the assumption that among nonlymphoid tumors glioblastoma cells are at the present time unique for their capacity to release immunosuppressive factors in vivo and in vitro.

Depression of immune responsiveness has been documented in patients with glioblastoma (1-6). Impaired cell-mediated immunity in glioblastoma patients is indicated by depressed skin reactivity to common antigens, decreased ability to become sensitized to dinitrochlorobenzene, and decreased in vitro lymphocyte responsiveness to PHA or antigens. Factors released by the tumor cells may well account for the described T cell immune deficiency state. This idea is supported by the detection of inhibitory factors on T cell activation present in glioblastoma culture SN in vitro and the cyst fluid of glioblastoma tumors in vivo. In addition the sera of glioblastoma patients contain a factor that inhibits the mixed lymphocyte reaction and the PHA- and Con A-induced lymphocyte proliferation in vitro. After removal of the glioblastoma by neurosurgery, suppressor activity in the sera disappeared (1, 4).

Although it is well documented that in most glioblastoma patients, humoral immune responses to their tumors develop (23, 24), there is little evidence of significant cellular anti-tumor immune responses (25,-27). In general, cytotoxic T cells have been implicated to contribute to the development of the cellular anti-tumor immune reactions (28, 29). The IL 2 dependency of the generation of CTL has been demonstrated by in vivo administration of highly purified IL 2 in concert with injections of tumor cells, which resulted in an augmented CTL and natural killer cell response in mice (30). The observed blocking of the generation of CTL by the glioblastoma cell-derived 97.000-m.w. factor according to the invention may account at least in part for inefficient immunosurveillance against glioblastomas. In

Impaired host immunocompetence may also emanate from lymphokine-induced production of mucopolysaccharide coats by glioblastoma cells that nonspecifically suppress the cellular immune response (31).

In addition to its relevance for modulation of T cell activation the supernatant containing the factor is of interest for its neuroblast growth inhibitory factor (NGIF)-like activity. NGIF is released by fetal rat glioblasts and exerts suppressor activities on neuroblasts but not on fibroblasts (31, 32). In addition, this 75.000-m.w. factor possessed the activity to promote morphological differentiation of Neuro 2A cells as evidenced by the formation of neural processes (32, 33). In an analogous assay we also observed that the Sephacryl S-200 glioblastoma fractions, which inhibited T cell responsiveness to IL 2 and the growth of neuroblasts, did induce neurile outgrowth.

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The following examples illustrate the invention. The following abbreviations are employed:

- IL 1, Interleukin 1; IL 2, Interleukin 2; NGIF, neuroblast growth inhibition factor; SN, supernatant; GFA, glial fibrillary acidic protein; DMEM, Dulbecco's modified Eagle's medium; ConA, concanavalin A; 2-ME, 2-mercapto-ethanol;  $^3\text{H}$ -Tdr. [ $^3\text{H}$ ]thymidine; PHA, phytohemagglutinin; OVA cells, ovalbumin-specific T-cell line; CTL, cytotoxic T-lymphocytes; MLC mixed lymphocyte culture; LPS, Lipopolysaccharide; AED, N-iodoacetyl-N-[5-sulfonic-1-naphthyl]ethylene diamine; CH, cycloheximide.

#### Example 1

- 10 Four human glioblastoma cell lines are established and maintained in mono-layer cultures by the method described in Diserens et.al., Acta Neuropathol. 53:21. Four cell lines used in the examples are designated C1-18; C1-229; 992 and 308 and were maintained for 72, 28, 8 and 4 months respectively.
- 15 Characterization of the glioblastoma cell lines revealed that cells from C1-229, 308 and 992 expressed Ia-like antigens on their surface, cells from 308 and 992 were positive for glial fibrillary acidic protein (GFA), and cells from C1-18, 308 and 992 expressed a neuro-ectodermal antigen identified by recently described monoclonal antibodies
- 20 (Acta Neuropathol. 57:158 Carrell S., et.al.)

For production of SN the glioblastoma cells were plated at  $1 \times 10^4$  to  $1 \times 10^6$  cells/well ( $2.4 \times 1.7$  cm; Linbro Scientific) in culture medium consisting of Dulbecco's modified Eagle's medium (DMEM), 10% fetal calf serum (FCS), and 300 ug/ml L-glutamine. Twenty-four hours after plating

25 the glioblastoma cells, the medium was replaced with 0.5 ml culture medium. The SN were collected 1 to 5 days later, centrifuged ( $2000 \times G$ , 10 min) and ultrafiltrated by using a micropartition system with YMS membrane (MPS.1, Amicon). The material deposited after centrifugation ( $1500 \times G$ , 15 min) on the membrane (m.w.  $> 10,000$ ), was

30 resuspended in RPMI 1640 to give the original volume and passed through  $0.45\text{-}\mu\text{m}$  filters.

Example 2 : Thymocyte proliferation assay

The effect of glioblastoma SN on the thymocyte proliferative response to concanavalin A (Con A) was tested in the following way. Samples of 50  $\mu$ l of SN at various dilutions were added to  $6 \times 10^5$  thymocytes from C<sub>3</sub>H/HeJ mice: thymocytes were suspended in 150  $\mu$ l of RPMI 1640 medium supplemented with 300  $\mu$ g/ml L-glutamine,  $1 \times 10^{-5}$  M 2-mercaptoethanol (2-ME), and 5% FCS in flat-bottomed microtiter plates and incubated for 72 hours in the presence of Con A (1  $\mu$ g/well). Sixteen hours before harvest 0.5  $\mu$ Ci of <sup>3</sup>H-Tdr, 5.0 Ci/mmol) was added per well.

10 The results are shown in Figures 1A and B as percent suppression compared with the Con A response of thymocytes treated with a medium control.

In Figure 1A the SN of the four glioblastoma cell lines used were harvested after 1, 3 and 5 days and tested at a final dilution of 1/10 on the thymocytes. In Figure 1B the testing took place on SN of 308 cells  
15 harvested after culture for 5 days at different seeding densities and final dilutions. In the absence of glioblastoma SN or medium control the background count was  $463 \pm 229$  and the Con A response  $76.797 \pm 6.631$  cpm.

The magnitude of inhibitory potency of the SN can be seen to be dependent on time of culture and cell density.

20 In selected experiments on metabolic inhibition using 308 cells the production of SN was performed with irradiated (2000 R) or mitomycin C-treated (50  $\mu$ g/ml, 30 min, 37°C) glioblastoma cells. Alternatively cycloheximide ( $10^{-5}$  M) was added to glioblastoma cells at the beginning of the 72-hour culture; cycloheximide was also added to control SN  
25 collected after 72 hours. These SN were ultrafiltrated three times before testing to remove CH.

The results of these experiments are summarised in Figure 2, where the SN of  $10^6$  cells cultured over 5 days were employed. As a CH control the SN of CH treated 308 cells and of 308 SN supplemented with CH after collection were untrafiltered.

- 5 In a further control, experiments were carried out on four established human tumor cell lines unrelated to glioblastoma cells. These results are summarised in Table 1.

TABLE 1

Effects of SN from various tumor cell lines on lymphocyte blastogenesis

10	Type of Malignancy <sup>a</sup>	Designation	Con A Response <sup>b</sup> (cpm $\pm$ SD)	% Suppression
	Neuroblastoma	SR.N.SH	52.123 $\pm$ 1.817	6.0
	Neuroblastoma	IMR.32	50.041 $\pm$ 2.751	9.2
	Melanoma	Me.43	58.382 $\pm$ 447	0
15	Rhabdomyosarcoma	RD	51.720 $\pm$ 904	6.8

<sup>a</sup> Tumor cells were cultured at a seeding density of  $10^6$  cells/ml for 5 days.

In the presence of Con A, thymocytes were incubated with 1/10 dilution of tumor cell SN or a control culture medium for the tumor cell SN. In the presence of medium control the background count of thymocytes was  $432 \pm 112$  and the Con A response was  $55441 \pm 4817$ .

20

It will be noted that the release of immunosuppressive factors does not appear to be a general property of cultured tumor cells.

Analysis of the L-arginine content in 308 SN revealed that inhibition is not due to arginine degradation in the medium by an arginase release by 308 cells. Furthermore, the 308-mediated inhibition could not be explained by the production of interferon as no antiviral activity was  
5 detected in 308 SN.

Example 3 Effect of glioblastoma SN on B lymphocytes and non-lymphoid cells

Mouse fibroblasts (A9, 3T3, LS cell lines) were cultured in DMEM supplemented with L-glutamine (300 ug/ml) and 10% FCS: mouse neuroblasts (Neuro 2A, NB4, 1A3 cell lines) were plated in Earle's minimum essential  
10 medium with 10% FCS. L-glutamine (300 ug/ml), and 1% nonessential amino acids (100x). Fibroblasts and neuroblasts ( $10^4$  cells/well) were incubated in 0.2 ml of medium for 72 hours. For the final 6 hours, the cells were pulsed with 1.2 uCi of  $^3\text{H}$ -Tdr. Thereafter, the medium was aspirated from each well, replaced with trypsin-EDTA. and incubated ( $37^\circ\text{C}$ , 10 min).  
15 Finally, the cells were harvested with the use of an automated harvester.

The data in Table II indicate that when compared with its effect on thymocytes, the 308 SN was less inhibitory on B cell proliferation: the percentage of inhibition of the LPS-induced stimulation of spleen cells was 29%, whereas the corresponding value for the myeloma cell line  
20 X63-Ag8 was 21%.

There was no difference in thymidine uptake between fibroblast cultures containing 308 SN and those being supplemented with a medium control. This was true for all three fibroblast cell lines tested. In contrast, the glioblastoma SN significantly suppressed the growth of  
25 the two neuroblast cell lines as measured by the uptake of  $^3\text{H}$ -Tdr (Fig.3, Table II) and by counting the number of cells at termination of the 72 hr cultures. Twenty-four, 48 and 72 hr after culturing  $1 \times 10^4$  Neuro 2A cells/well, the cell numbers  $\times 10^4$ /well of quadruplicate cultures were in the presence of the 308 SN:  $9.8 \pm 0.17$  (24 hr),  $1.80 \pm 0.36$  (48 hr), and  
30  $4.3 \pm 0.44$  (72 hr), whereas the corresponding values for the medium controls were  $0.88 \pm 0.05$ ,  $2.55 \pm 0.06$ , and  $6.08 \pm 0.40$ , respectively.

TABLE II

Effect of 308 SN on lymphoid and nonlymphoid cells

Cell Type	Description or Strain	Stimulant	<sup>3</sup> H-Tdr Uptake(cpm)		
			Medium control	308 SN <sup>a</sup>	% Suppression
Thymocytes	C3H/HeJ	Con A	66.709 ± 3.104	8.323 ± 235	88
T cell lymphoma	EL-4	-	137.960 ± 25.173	164.487 ± 6.253	0
Spleen cells	DBA	LPS <sup>b</sup>	72.442 ± 3.886	50.990 ± 2.184	29
Myeloma cells	X63-Ag8 <sup>b</sup>	-	117.090 ± 2.597	92.580 ± 6.351	21
Neuroblasts <sup>c</sup>	NB41A3	-	48.073 ± 1.756	15.504 ± 833	68
Neuroblasts	Neuro 2A	-	250.807 ± 19.700	62.403 ± 6.108	76
Fibroblasts <sup>c</sup>	A9	-	78.591 ± 11.029	78.323 ± 10.444	0
Fibroblasts	3T3	-	117.316 ± 8.484	117.011 ± 4.026	0
Fibroblasts	L929	-	84.395 ± 6.189	92.391 ± 5.674	0

<sup>a</sup> All assays were performed with the same 308 SN added to give a final dilution of 1/10.

<sup>b</sup> After lysis of erythrocytes  $6 \times 10^5$  spleen cells were stimulated with lipopolysaccharide (LPS, E. coli 0127 B8, Difco Lab.). The mouse X63-Ag8 myeloma cells were cultured at  $10^4$  cells/well in DMEM supplemented with 10% FCS.

<sup>c</sup> See legend of Figure 3.

In Figure 3 the SN of  $10^6$  cells cultured for 5 days were employed together with a medium control.

#### Example 4 Mechanisms of glioblastoma SN-induced inhibition of T-cell activation

Thymocytes were treated with 308 SN (final dilution of 1/10) in the presence of PHA (0.5 ug/well) and various dilutions (1/10 in 1/160) of IL 1 or IL 2. In the absence of interleukins the background count was 209 ± 84 and the PHA response was 3074 ± 425.

The IL 1 preparation used was prepared from LPS-stimulated peritoneal cells with Con A-Sepharose as described in reference 7.

The results are summarised in Table III.

TABLE III

Effect of 308 SN on thymocytes stimulated with PHA and various doses of IL 1 or IL 2

5

Effect of 308 SN on thymocytes stimulated with PHA and various doses of IL 1 or IL 2

Stimulant <sup>a</sup>		<sup>3</sup> H-Tdr Uptake (cpm)		% Suppression
		Medium control	308 SN	
10	PHA + IL 1 <sup>b</sup>			
	1/10	76.342 ± 5.978	12.104 ± 612	84.2
	1/20	69.631 ± 5.150	13.423 ± 212	81.8
	1/40	42.143 ± 1.706	9.061 ± 842	78.5
	1/80	18.407 ± 128	2.648 ± 221	85.7
15	PHA + IL 2 <sup>b</sup>			
	1/10	84.457 ± 3.621	13.551 ± 817	84.0
	1/20	84.882 ± 4.185	15.492 ± 1.005	81.8
	1/40	76.537 ± 2.404	11.250 ± 1.423	85.4
	1/80	43.244 ± 1.753	5.973 ± 175	86.2
	1/160	26.505 ± 1.044	3.182 ± 3.279	88.0

<sup>a</sup> Thymocytes were treated with 308 SN (final dilution of 1/10) in the presence of PHA (0.5 µg/well) and various dilutions (1/10 to 1/160) of IL 1 or IL 2. In the absence of interleukins the background count was 209 ± 84 and the PHA response was 3074 ± 425.

<sup>b</sup> The IL 1 preparation used was prepared from LPS-stimulated peritoneal cells of BALB/c mice; the IL 2 was prepared by stimulation of BALB/c spleen cells with Con A-Sepharose as described previously (17).

These results show that inhibition of the Con A response of thymocytes apparently results from influences on late events of the T cell activation cascade, because the 308 SN inhibited the proliferative response of thymocytes stimulated with PHA in the presence of IL 1 or IL 2. The degree of 308 SN-induced suppression of proliferation was similar at various concentrations of IL 1 or IL 2 employed.

EXAMPLE 5:Effect of SN-308 on cloned T-cell lines

Proliferation of H-2<sup>b</sup>-restricted hapten (N-iodoacetyl-N-(5-sulfonic-1-naphthyl) ethylene diamine [AED])-specific cloned cytotoxic T cells (CTL) (10) was assessed in culture medium consisting of Iscove's modified DMEM supplemented with 10 % FCS,  $1 \times 10^5$  M 2-ME, anti-  
5 bodies, and 10 % (v/v) of a 24-hr spleen cell Con A SN (IL 2 SN). The CTL ( $10^4$ /well) were cultured alone or together with  $10^6$  irradiated (2000 rad) haptenated spleen cells (H-2<sup>b</sup>) as stimulators. Various dilutions of SN of 308 glioblastoma cells or of a control  
10 medium were added to the mixture of the CTL and stimulators cultured in culture medium for 72 hr. Sixteen hours before harvest, each well was pulsed with 1.2  $\mu$ Ci of <sup>3</sup>H-Tdr.

308 glioblastoma SN was also tested on cloned ovalbumin-specific T cells (OVA cells). The OVA cells were cultured for 48 hr in  
15 0.2 ml culture medium (see above) with IL 2 SN. The IL 2-dependent proliferation was tested in the presence of various dilutions of 308 SN or medium control. The incorporation of <sup>3</sup>H Tdr (1.2  $\mu$ l/well) was measured over a 12 hr period.

The results are shown in figures 4A and 4B. In A. hapten-specific  
20 cloned CTL were cultured (at  $10^4$  cells) alone (R) or together with  $10^6$  irradiated (2000 rad) AED-haptenated spleen cells as stimulators (S) (black bar). Various dilutions (1/8 to 1/128) of 308 glioblastoma SN or a 1/8 dilution of control medium (.....) were added to the mixture of CTL and cultured for 72 hr. In B  $10^4$   
25 OVA cells were cultured for 48 hr with IL 2 SN (black bar) or without IL 2 SN (——). The 308 glioblastoma SN (1/8 to 1/128 diluted) or a 1/8 dilution of control medium (.....) was added to OVA cells in the presence of IL 2 (final dilution 1/20). Data are given as counts per minute of <sup>3</sup>H-Tdr incorporation.



Figure 4A shows that cells of an H-2-restricted, hapten-specific T cell line proliferated in the presence of IL 2 and haptenated, irradiated spleen cells as stimulator cells. This proliferation was completely blocked by adding the glioblastoma-produced factor. Maximal inhibition was noted when the factor was added in OVA T cells growing in the presence of IL 2 only (Fig. 4B).

From the experiments outlined above, the inhibition of T cell growth by glioblastoma cell-derived factors could be due to their direct interference with IL 2-triggered events leading to T cell proliferation. This possibility is supported further by the finding that the 308 SN had no effect on the growth of a thymoma cell line that grows of a thymoma cell line that grows independently of IL 2 (Table II).

#### EXAMPLE 6:

The release of analogous inhibitory factors by the glioblastoma cells in vivo was demonstrated by testing the ultrafiltrated cyst fluid of the patient from whom the 308 cells originated. The cyst fluid (final dilution 1/20), ultrafiltrated on YMB membranes, induced a 28% inhibition of the IL 2-induced proliferation of OVA T cells, whereas 12 control sera had no effect.

#### EXAMPLE 7:

##### Effect of glioblastoma SN on the induction of cytotoxic T-cells

Anti-H-2<sup>b</sup>-specific CTL were induced by stimulation of  $12 \times 10^6$  B10.D2 (H-2<sup>d</sup>) spleen cells with  $30 \times 10^6$  irradiated (2000 R) C57B1/6 (H-2<sup>b</sup>) stimulator spleen cells in 6 ml Iscove's modified DMEM containing 10 % FCS,  $1 \times 10^{-5}$  M 2-ME, and antibiotics (mixed lymphocyte culture (MLC) medium). After 6 days the specific cytotoxic activity of the CTL was tested in a 3-hr <sup>51</sup>Cr-release

assay in 0.2 ml cultures containing  $10^4$   $^{51}\text{Cr}$ -labeled target cells EL-4 ( $\text{H-2}^b$ ), P815 ( $\text{H-2}^d$ ), Yac-1 ( $\text{H-2}^d$ ) at different target ratios as described in 11. The cytotoxic effect of the cells cultured over 6 days in MLC medium was compared with the effects of cells cultured in MLC medium being supplemented with medium control or 308 SN.

The results are summarized in Fig. 5A-D. Legend Fig. 5.

The cytotoxic effect of the cells cultured over 6 days in MLC medium (O-O) was compared with the effects of cells cultured in MLC medium being supplemented with a) medium control (final dilution 1/10) ( $\square$ — $\square$ ), b) 308 SN ( $\bullet$ — $\bullet$ ) diluted 1/10 (A-D), 1/100 or 1/1000 (A) or with c) Sephacryl S-200 column fractions that were known to inhibit ( $\blacktriangle$ — $\blacktriangle$ ) or to have no effect ( $\triangle$ — $\triangle$ ) on the IL 2-dependent proliferation of OVA T cells. The fractions to be assayed were concentrated 10 x on YMB membranes, and the concentrated fractions were added to MLC medium to give a final dilution of 1/10. Results are given as percent specific  $^{51}\text{Cr}$  release (see Reference 11). Test duration 3 hr. Mean values of three samples.

The preparation and testing of Sephacryl S-200 column fractions is described below).

As shown in figure 6 the addition of IL 2 together with the 308 SN factor did not restore the generation of CTL. A 2-hr preincubation of mature CTL with 308 SN had no influence on the function of CTL on their targets.

In Fig. 6 A during the entire culture period of 6 days, IL 2 (final concentration of 10 % Con A SN) was present in MLC treated with 308 SN (1/10 diluted) ( $\bullet$ — $\bullet$ ) or medium control ( $\circ$ — $\circ$ ). After 6 days the CTL activity was tested on EL-4 cells. In B. before testing on EL-4 cells, CTL generated over 6 days in culture medium

were preincubated for 2 hr in medium control or 308 SN (1/4 diluted).

EXAMPLE 8:

Biochemical characterisation of the glioblastoma SN

The 308 glioblastoma SN obtained from 308 cells cultured in serum  
5 free conditions over 96hr was concentrated 30-fold by ultra-  
filtration on YM-1C (Amteon) membrane, and 1.5 ml of concentrated  
SN was applied on a Sephacryl S-200 (column size 2.5 x 85 cm) at  
4°C as described (9). The eluate collected in 2.5 ml fractions was  
monitored for adsorbance at 280 nm and was tested on fibroblasts  
10 (IL 929) and neuroblasts (Neuro 2 A) as described above. In  
addition the fractions were tested on OVA cells in the presence of  
IL2 and on thymocytes stimulated with PHA. The column, in a parallel  
run, was calibrated with  $\gamma$ -globulin ( $\gamma$ -Glob. m.w. 150.000), bovine  
serum albumin (BSA. m.w. 67.000), ovalbumin (OVA, m.w. 45.000),  
15  $\alpha$ -chymotrypsinogen (CHY. m.w. 25.000), and cytochrome c (CYT C.  
m.w. 12.400).

These results are summarized in figure 7 whereby the upper table  
shows the calibration run. Results are given as stimulation index  
(SI) comparing proliferation of cells treated with column fractions  
20 to cells treated with medium control. Results of serum-free control  
medium fractionated on a Sephacryl S-200 column are shown on OVA  
cells treated with IL 2 ( $\square$ — $\square$ ).

The peak of the inhibitory activity on neuroblasts and OVA cells  
was eluted from the column at the identical position, with an  
25 apparent m.w. of 97.000 (Fig. 7). Sephacryl S-200 fractions of  
a medium control consisting of DMEM (Fig. 7) and a medium control  
supplemented with 10 % FCS (data not shown) had no inhibitory  
effects on the growth of OVA cells stimulated with IL 2. Further-

more, no inhibitory effect was observed when testing 308 SN fractions on fibroblasts. In fact, the growth of fibroblasts but not of neuroblasts or of IL 2-treated OVA cells was augmented by fractions that exhibit an apparent m.w. of around 22,000. These  
 5 fractions were identified to additionally stimulate the PHA response of mouse thymocytes (Fig. 7) but to have no effect on the growth of OVA cells cultured in the absence of IL 2: the incorporation of  $^3\text{H}$ -Tdr by OVA cells was  $284 \pm 72$  and in the presence of IL 2 (final dilution 1/10) was  $62038 \pm 2931$ . When OVA cells were  
 10 treated with 308 fractions eluting from the column between m.w. 150,000 and 10,000, no proliferation of the OVA cells was observed, with cpm values never exceeding 500. This indicates the absence of IL 2 in 308 fractions including those that were found to augment the PHA response of thymocytes.

#### EXAMPLE 9:

15 Pools of chromatographed fractions of the 308 SN were assayed for their ability to enhance the release of IL 2 by spleen cells treated with suboptimal doses of Con A. and the results summarized in Fig. 8.

In Fig. 8 A fractions were assayed for their capacity to augment the PHA response of thymocytes: data are given as stimulation  
 20 index (SI) comparing the PHA response of thymocytes treated with column fractions to untreated thymocytes. EF, effluent volume (ml). In B, pooled fractions (pool I to V) were concentrated 10 x on YMB membranes and were tested for their ability to enhance the release of IL 2 by spleen cells ( $5 \times 10^6/\text{ml}$ ). After 44 hr the spleen cell SN  
 25 were harvested, and the IL 2 activity was assayed on OVA cells. The activity in test samples was transformed into units as described in reference 13.

#### Activity

=  $\frac{\text{Reciprocal titer of test sample at 30 \% of maximal standard cpm}}{\text{Reciprocal titer at 30 \% of maximal standard cpm}}$   
 30

In the absence of pooled fractions of 308 SN the amount of IL 2 released by spleen cells treated with Con A ( $0.5 \mu\text{g}/\text{ml}$ ) was 21 U/ml: upon treatment with optimal doses of Con A ( $5 \mu\text{g}/\text{ml}$ ) the IL 2 content measured was 97 U/ml.

- 5 Compared with Con A alone, pool I to III neither stimulates nor inhibited the release of IL 2 (Fig. 8). However, with fraction pool IV and V the increase of the amount of IL 2 released was 3.5- and 1.6-fold, respectively (Fig. 8). Thus the capacity to enhance the release of IL 2 by spleen cells appears to be  
10 associated with fractions augmenting the PHA response of thymocytes.

- In addition to inhibiting neuroblast growth, the 308 fractions were found to induce the outgrowth of neurites of Neuro 2A cells: 66 % of Neuro 2A cells treated with fraction 93 (peak inhibitory fraction on neuroblast growth) showed cell processes, whereas  
15 the corresponding values in cultures treated with control fractions (fraction 112) or control medium were 21 and 22 %, respectively.

5 The immunosuppressant factor according to the invention is indicated for use in the treatment of diseases and conditions where suppression of the body's immune response is desired. Thus use in connection with transplant operations to prevent rejection is indicated. Examples of diseases where suppressions of the body's immune systems is indicated include the so-called auto-immune diseases.

10 The factor according to the invention can be employed in mixed form as purified supernatant from appropriate glioblastoma cell-lines or in fractionated and purified form as hereinbefore described.

It can be set up in suitable form for administration in a manner conventional for substances of this nature in accordance with the condition to be treated.

CLAIMS:

1. An immunosuppressant factor derived from human glioblastoma cells substantially as hereinbefore characterised.
2. A factor as claimed in claim 1 which inhibits interleukin 2 dependant T-cell mechanisms.
3. A factor as claimed in claim 1 having an approximate molecular weight of 97,000.
4. A factor for the inhibition of neuroblast growth substantially as hereinbefore characterised.
5. A factor as claimed in claim 4 having an approximate molecular weight of 75,000.
6. An interleukin 1 like factor derived from human glioblastoma cells substantially as hereinbefore characterised.
7. A factor as claimed in claim 7 having an approximate molecular weight of 22,000.
8. A supernatant harvested from cultured human glioblastoma cells containing a factor according to any one of claims 1 to 3 and/or claims 4 and 5 and/or claims 6 and 7.
9. The steps, features, compositions and compounds referred to or indicated in the specification and/or claims of this application, individually or collectively, and any and all combinations or any two or more of said steps or features.

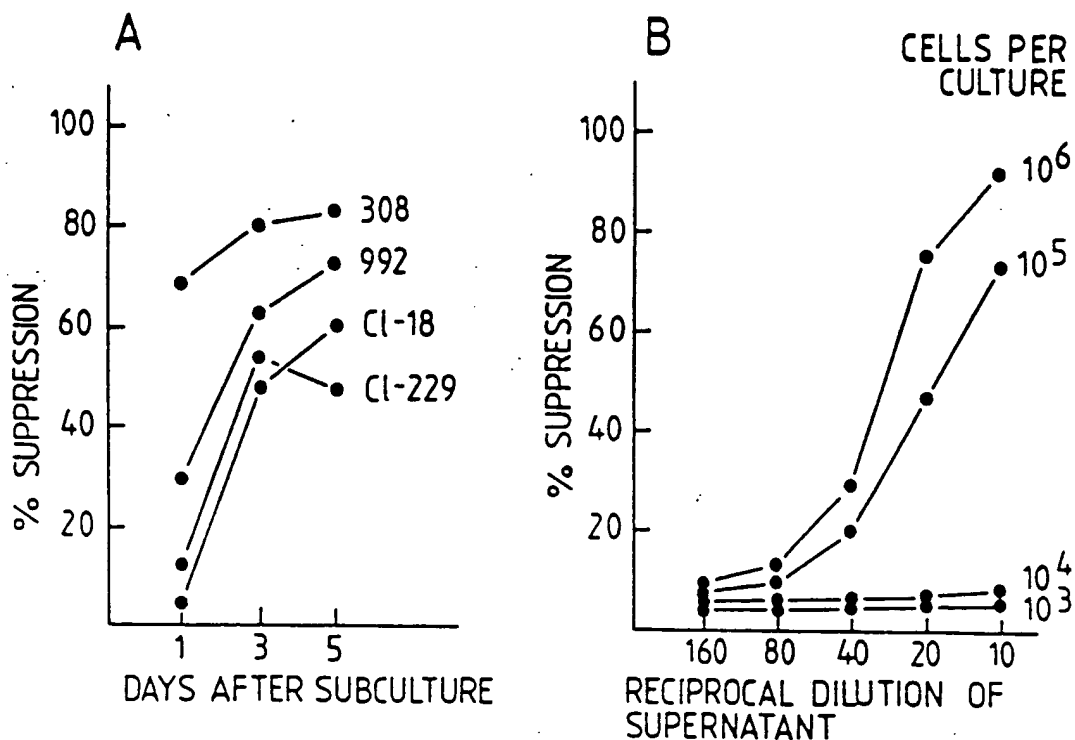


FIG. 1

TREATMENT OF  
308 CELLS

—

MITOMYCIN C

2000 RAD

CYCLOHEXIMIDE  
 $10^{-5}M$

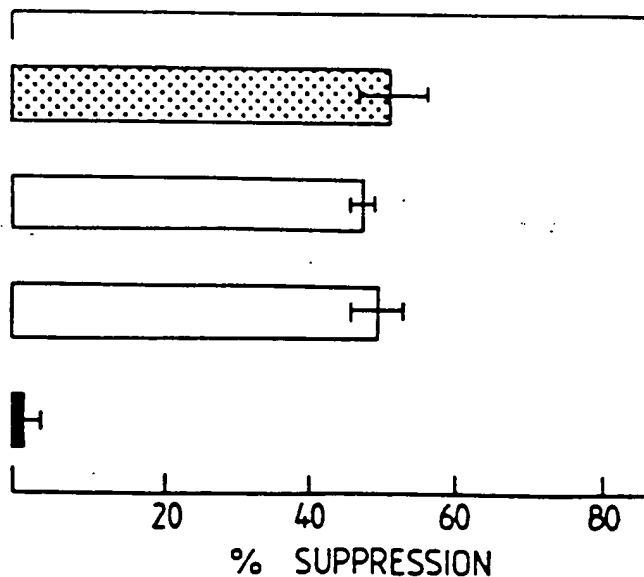


FIG. 2



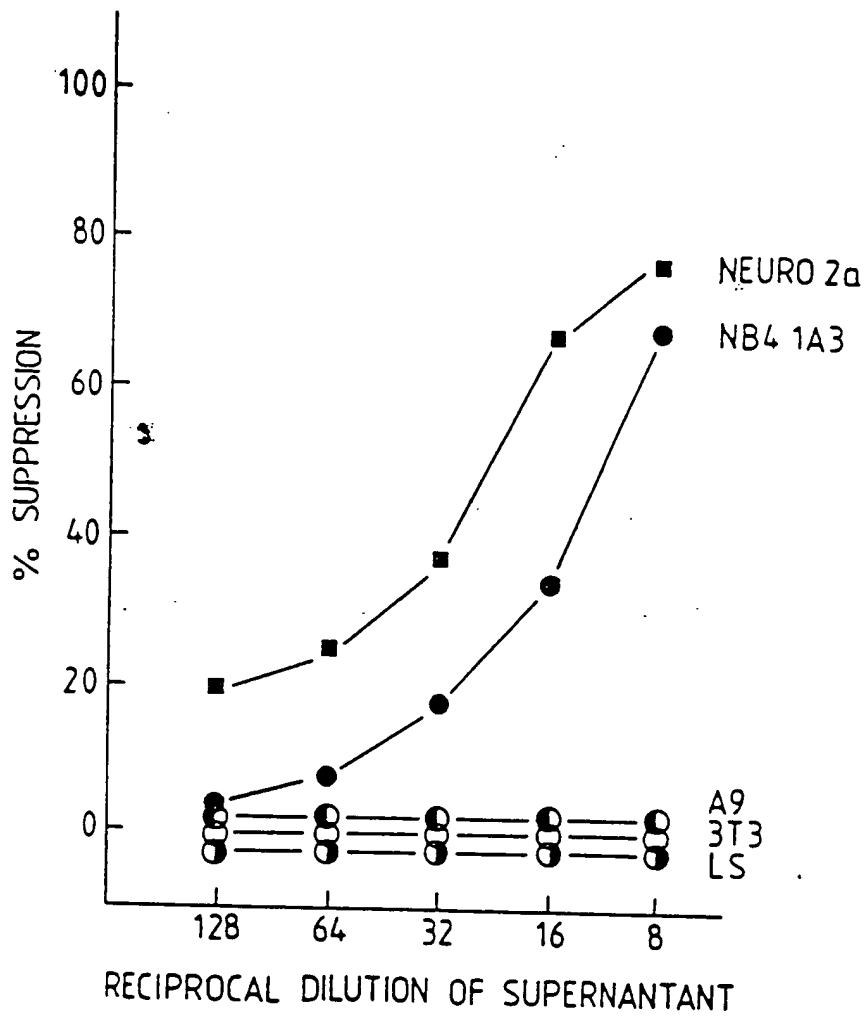


FIG. 3

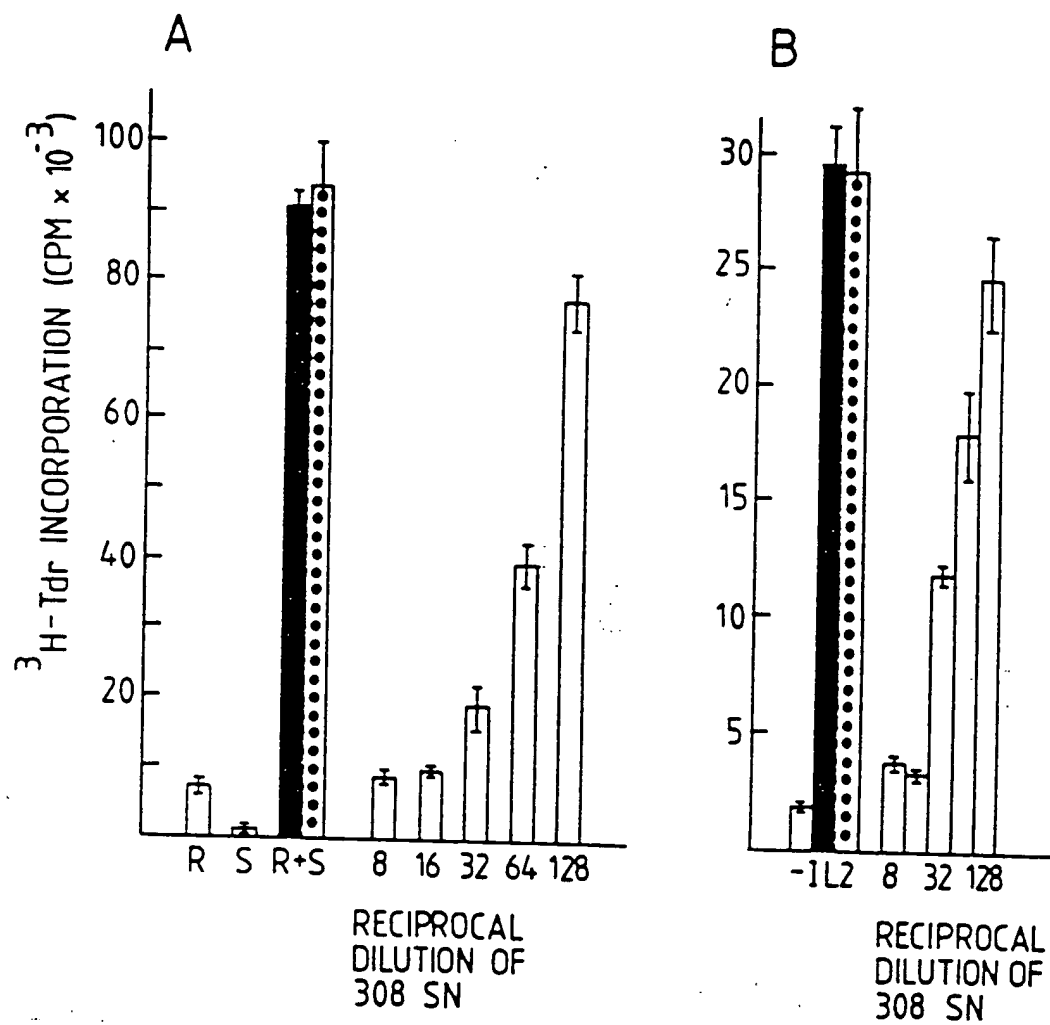


FIG. 4

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Case 0155433

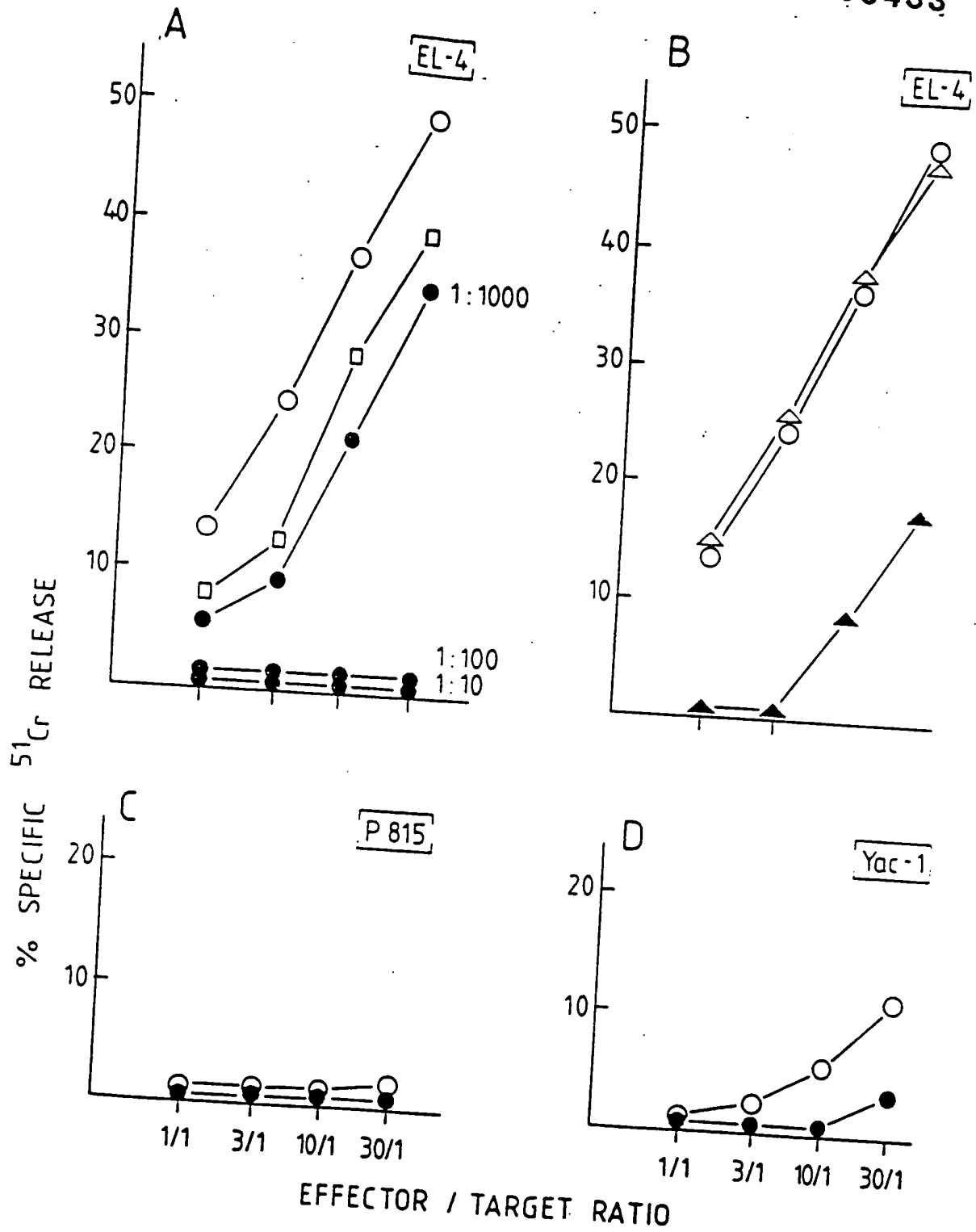


FIG. 5

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0155433  
Case 118-6323

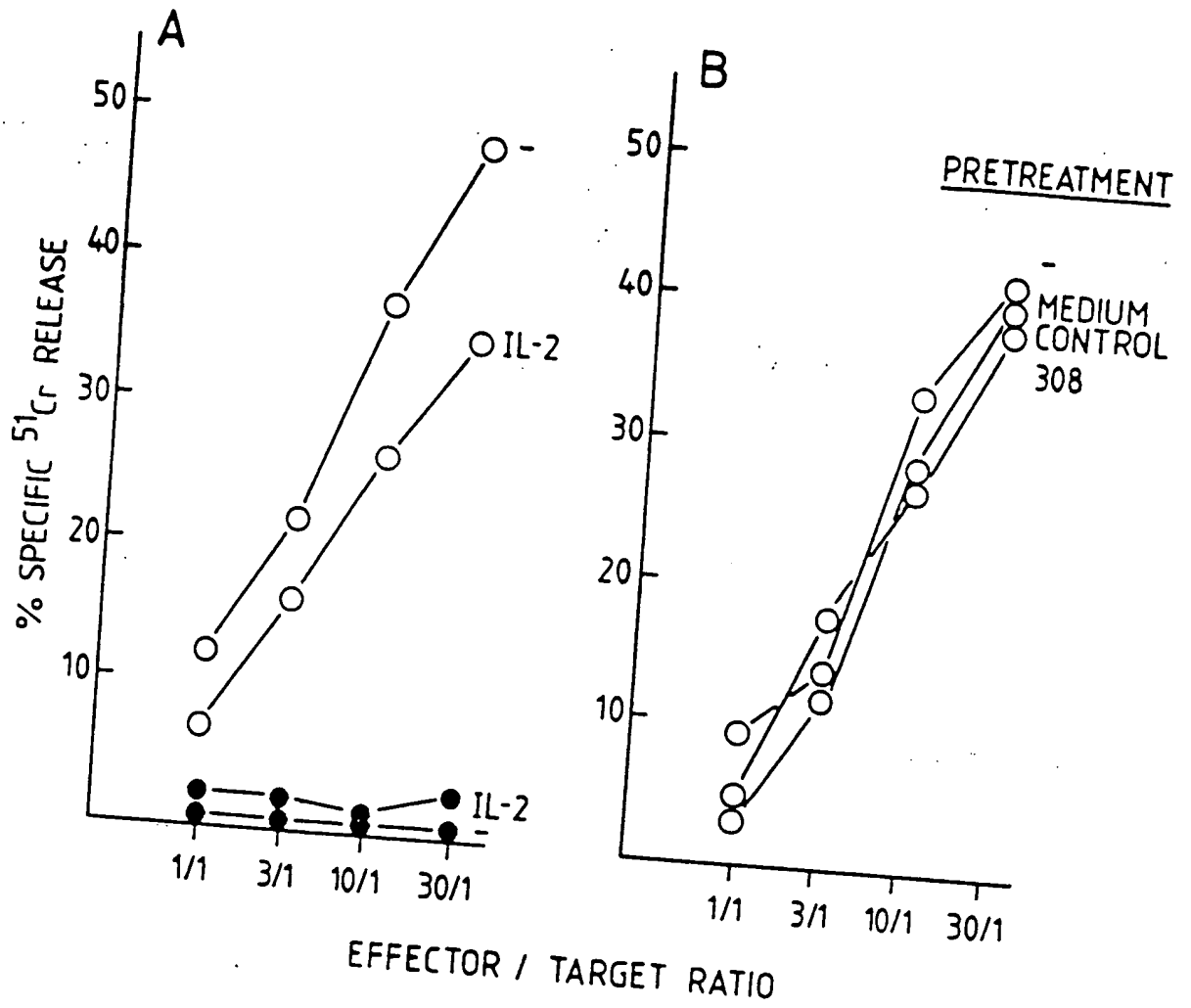


FIG. 6

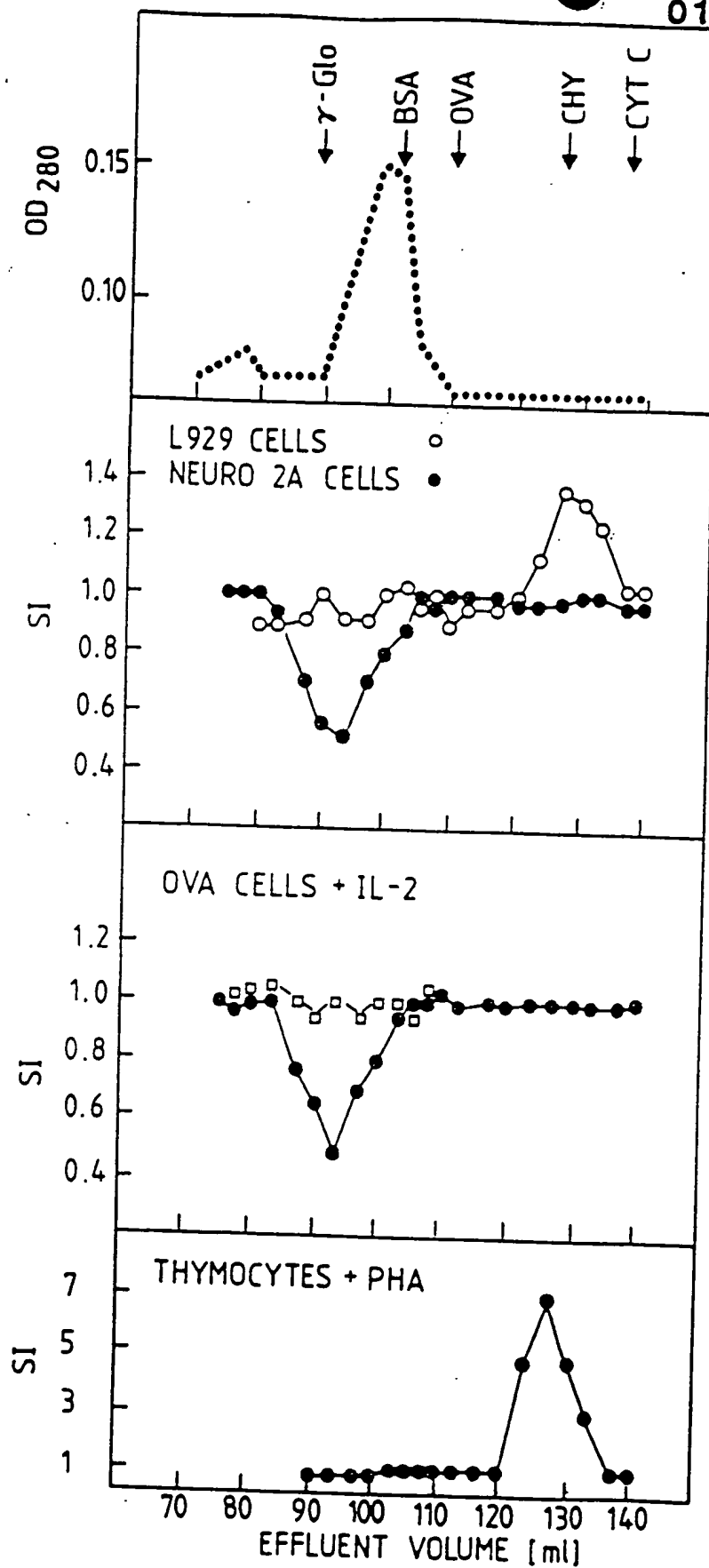


FIG. 7

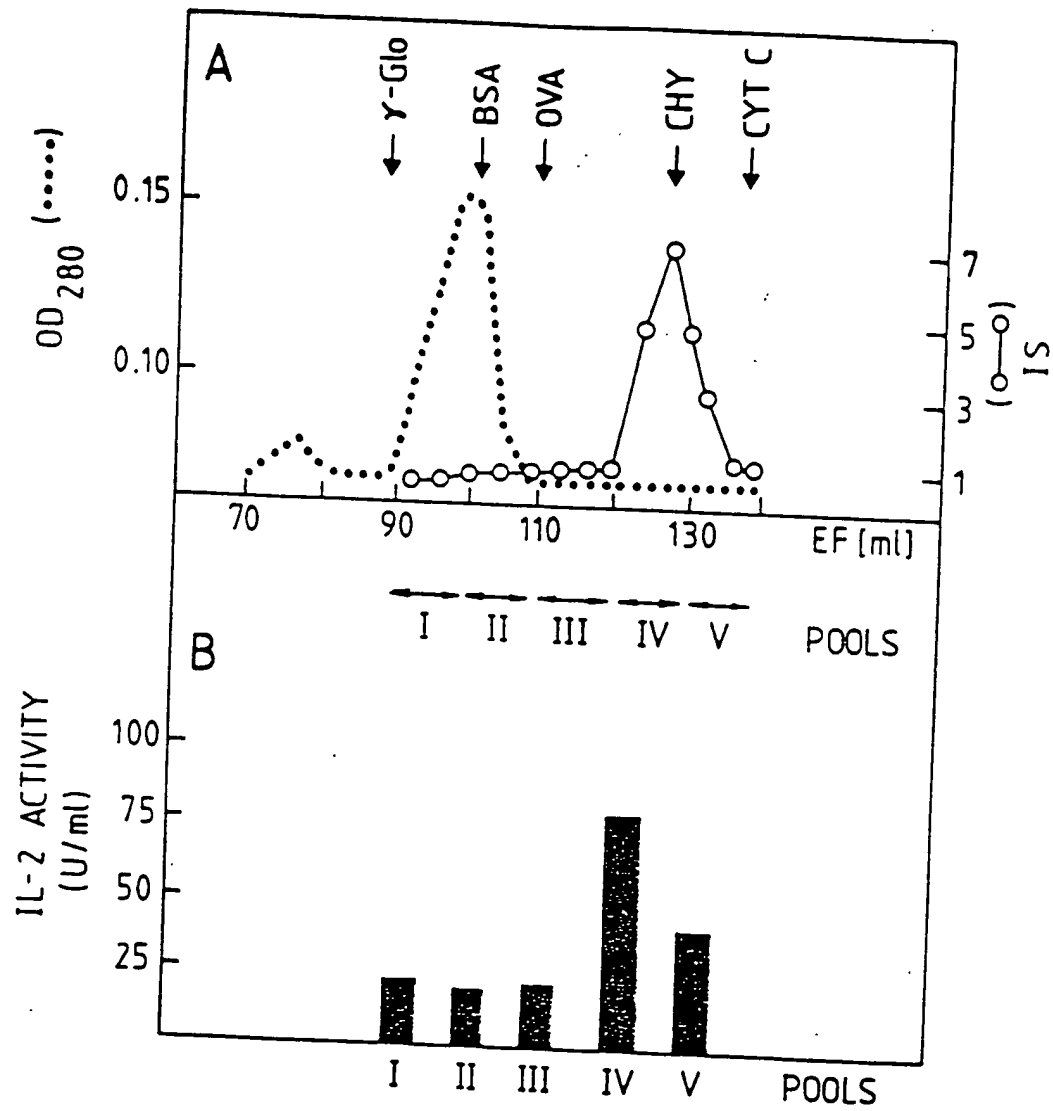


FIG. 8



DOCUMENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int. Cl. 4)
X	BIOLOGICAL ABSTRACTS/RRM, vol. 27, 1984, abstract no. 45949, Philadelphia, US; A. FONTANA et al.: "Immunodeficiency in glioblastoma patients, glioblastoma cells release factors inhibiting interleukin 2 mediated effects on T cells" & NEUROLOGY (USA) 1984, vol. 34, no. 3, suppl. 1, page 184 * title and "terms" *	1-3	C 12 P 21/00 A 61 K 37/02 // (C 12 P 21/00 C 12 R 1/91 )
D,A	--- THE JOURNAL OF IMMUNOLOGY, vol. 129, no. 6, december 1982, pages 2413-2419, US; A. FONTANA et al.: "Production of prostaglandin E and on interleukin-1 like factor by cultured astrocytes and C6 glioma cells" * whole document *	1-9	TECHNICAL FIELDS SEARCHED (Int. Cl. 4)  C 12 P C 12 N A 61 K
The present search report has been drawn up for all claims			
Place of search THE HAGUE		Date of completion of the search 06-12-1984	Examiner RYCKEBOSCH A.O.A.
<b>CATEGORY OF CITED DOCUMENTS</b> X : particularly relevant if taken alone Y : particularly relevant if combined with another document of the same category A : technological background O : non-written disclosure P : intermediate document T : theory or principle underlying the invention E : earlier patent document, but published on, or after the filing date D : document cited in the application L : document cited for other reasons & : member of the same patent family, corresponding document			